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Biliary cytology: a diagnostic tree for adenocarcinoma based on a cohort of 135 patients with Endoscopic Retrograde Cholangio-Pancreatography for stenosis of the extrahepatic bile duct

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3 1 **Biliary cytology: a diagnostic tree for adenocarcinoma based on a cohort of 135 patients**
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5 2 **with Endoscopic Retrograde Cholangio-Pancreatography for stenosis of the**
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7 **extrahepatic bile duct**
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12 5 **Short running title:** Diagnostic tree and atlas of bile
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40 17 **Precis for use in the Table of Contents**
41

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43
44 19 general cytological criteria.
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1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60

1 INTRODUCTION

2 Pancreatic ductal adenocarcinoma and cholangiocarcinoma of the biliary tract are the two
3 most common causes of malignant bile duct strictures, whose incidence has increased in the
4 last 30 years,¹ with a global incidence rate of 7.7 to 11 and 1.4 to 2.1 per 100,000 person-
5 years respectively.²

6 A suspicion of malignant stenosis of the extrahepatic bile duct leads patients to undergo
7 ERCP (Endoscopic Retrograde Cholangiopancreatography) for diagnostic purposes and, if
8 possible, therapy by the placement of a biliary prosthesis. The specificity of the cytological
9 examination of these biliary samples is remarkably high but reported sensitivities for the
10 diagnosis of malignancy are variable.³⁻⁶ As a result, cytological analysis of bile is very useful
11 to assess the diagnosis of pancreatobiliary carcinoma, enabling early and appropriate
12 management, without performing a new invasive procedure. However, the cytological
13 examination of bile is a challenge for the cytopathologist, who must be aware of diagnostic
14 pitfalls such as false positives (Primary Sclerosing Cholangitis (PSC), stent-related atypia) or
15 false negatives (samples with low cell-counts or very well-differentiated adenocarcinomas).⁷⁻⁹
16 Numerous cytological criteria have been proposed on biliary brush samples^{10,11} but, to our
17 knowledge, only three studies have performed multivariate analyses of malignant cytological
18 criteria on biliary aspiration samples, which are often less crushed and just as cellular as brush
19 specimens.^{12,15,16} These three studies, each performed by 3 experienced cytopathologists,
20 showed variable specificities (between 65 and 86.4%) and variable sensitivities (between 76.9
21 and 98%) and a proportion of well-differentiated adenocarcinomas of 43% and 63%
22 respectively for 2 out of the 3 studies.^{12,15} Since cytological examination of biliary aspiration
23 is not a common practice worldwide, our study aimed to define specific, sensitive and
24 reproducible cytological criteria for malignancy on bile aspiration samples by both a novice
25 and a 5 years experienced cytopathologist in biliary cytology and was designed for training

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3 1 purposes. Our first goal was to propose a diagnostic tree to help in diagnosis of extra-hepatic
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5 2 bile duct adenocarcinomas, and focused on the specificity rather than the sensitivity in order
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7 3 to avoid false positives. Our second goal was to elaborate an atlas illustrating the main studied
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9 4 cytological criteria and to assess its use in training novice cytopathologists in biliary cytology,
10
11 5 especially in well-differentiated adenocarcinomas.
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14 6 **MATERIALS AND METHODS**

15 7 **Patients and cytological samples**

16
17 8 Seventy two malignant biliary cytological samples were retrospectively collected from the
18
19 9 Grenoble Alpes University Hospital pathology department between January 1st, 2012 and
20
21 10 December 31st, 2018. All the biliary samples were obtained under ERCP for biliary stenosis
22
23 11 at the Grenoble Alpes University Hospital. From the 72 bile aspiration specimens available 3
24
25 12 were excluded as they were pauci-cellular and/or poorly preserved. The 69 malignant
26
27 13 retained samples came from pancreatic and extra-hepatic biliary tract adenocarcinomas all
28
29 14 proven by histology (8 pancreatoduodenectomy specimens, 45 pancreatic or biliary tract
30
31 15 punctures or 16 pancreatic metastasis). They corresponded to 56 pancreatic ductal
32
33 16 adenocarcinomas, 10 extra-hepatic cholangiocarcinomas, 1 adenosquamous carcinoma and 2
34
35 17 ampullary adenocarcinomas. Eighty one percent of them were well-differentiated, compared
36
37 18 to respectively 43% and 65% of well-differentiated adenocarcinomas in the 2 studies
38
39 19 previously mentioned ^{12,15}; 36 corresponded to men and 33 to women, ranging from 28 to 90
40
41 20 years old. This non-interventional, monocentric, retrospective study involving data and
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43 21 samples from human participants was carried out at the Grenoble University Hospital
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45 22 according to French current regulation and complies with Reference Methodology n°004
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47 23 issued by French Authorities (CNIL). Subjects were all informed and did not oppose; written
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49 24 consent for participation was not required for this study in accordance with the national
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51 25 legislation and the institutional requirements. The raw data supporting the conclusions of this
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1 article will be made available by the authors within respect of General Data Protection
2 Regulation, without undue reservation.

3 Supplementary biliary samples classified as benign (N=58) and atypical (N=11), with (N=49)
4 or without (N=20) a stent in place were retrospectively included for the second reading
5 between January 1st, 2009 and December 31st, 2014. None of them showed evidence of
6 malignancy during a follow-up of 5 years minimum (5-9 years). This prolonged follow-up
7 demonstrates that the atypical cytologies corresponded to reactive atypia and these 11 atypical
8 specimens were therefore considered as non-malignant for the statistical study. From these 69
9 selected non-malignant cases, 3 were excluded because of bad preservation or acellular slides.
10 The 66 retained cases corresponded to secondary stenosis after surgery for lithiasic disease
11 (N=58) or PSC (N=8).

12 **Collection of bile samples**

13 *Interventional endoscopy technique*

14 Patients with biliary stenosis underwent ERCP under general anesthesia on an X-ray table
15 using a video side-viewing duodenoscope positioned in the second duodenum facing the
16 papilla. A 0.035-inch-diameter guide-wire was introduced into the biliary tract and an
17 injection of contrast medium (Telebrix; Guebert; France) was performed. The opacification
18 objectified the presence of a biliary stenosis. After biliary sphincterotomy, a disposable
19 cytology brush was slipped over the guide-wire through the stenosis (Cook[®], France) with
20 several passes in the stenosis. Three to 10 mL of bile were collected half before and half after
21 brushing the stenosis. Brushing is performed before stenosis dilatation. Because the length of
22 the guide-wire tract at the distal end of the device was 6 cm, the entrance hole of the guide
23 wire had to be above the papilla to optimize bile aspiration. If required, the procedure was
24 completed by the installation of a temporary prosthesis pending the etiological diagnosis of
25 stenosis.

1 ***Cytological technique***

2 Each bile aspiration specimen before and after brushing was cooled on ice and sent to the
3 pathology department as soon as possible to avoid degenerative changes. Each specimen, one
4 bile before brushing and one bile after brushing, was centrifuged (2,000 revolutions per
5 minute for 10 minutes), and the deposit was smeared into 4 slides: 2 slides were stained with
6 Papanicolaou after using a Merckofix fixing spray (Merck France, Calais, France) and 2
7 slides were air-dried and stained with May Grünwald Giemsa (MGG).

8 ***Cytological examination***

9 For the first reading each of the 69 malignant samples was independently examined by a
10 cytopathologist novice in biliary cytology (AB) and by a 5 year-experienced cytopathologist
11 in biliary cytology (DG), both not blinded to the final diagnosis.

12 The presence or absence of 24 cytological criteria were studied, based on the *Papanicolaou*
13 *Society of Cytopathology* fascicle ⁷ and on prior studies on biliary brushings.^{10,11} These
14 criteria were divided in 5 categories (Table 1) corresponding to (1) the background of the
15 smears, (2) the presentation of the epithelial cells or clusters: “epithelial cell density” was
16 considered adequate if at least 5 clusters of cells were observed in each of the 8 slides ;
17 “nuclear overlap” was retained if moderate or marked ; “3D-cluster” was characterized by a
18 disorganized overlapping atypical cells cluster ; lumen within clusters characterized the
19 “acinar arrangement”; “single malignant cell” was defined as a single cell with intact
20 cytoplasm, enlarged nucleus occupying at least 50% of the cell area, chromatin changes,
21 irregular nuclear shape; “cytophagy” corresponded to an epithelial cell-in-cell arrangement,
22 (3) the cytomorphological aspect of the nuclei: “anisonucleosis” was noted if a size variation
23 of at least 4 times between the smallest and largest nuclei within a cell cluster was observed ;
24 “irregular nuclear shape”; “enhanced nuclear membrane” was noted if visible at a x20
25 objective lens ; increased N/C ratio (“N/C ratio > 0.5”) was noted when the nucleus occupied

1 more than 50% of the cell area ; “chromatin changes” included a clear, coarse chromatin or
2 hyperchromatism ; “prominent nucleolus” was noted when a nucleolus was visible at a x20
3 objective lens (4) the appearance of the cytoplasm : “vacuolated cytoplasm” ; “enhanced
4 cytoplasmic membrane” when the limits of the cytoplasm were too clearly visible, (5) and
5 two accompanying criteria such as “neutrophils phagocytosis” and the presence of
6 “degenerative cell”. The criteria from the categories (2) and (3) were analyzed in comparison
7 with the normal flaps of ductal cells arranged in a honeycomb pattern. At the end of the first
8 reading, an atlas (Fig. 1) was elaborated to better illustrate the cytological criteria studied,
9 especially those with lower inter-observer agreement.

10 The second reading was performed on the 69 malignant cases and on the 66 non-malignant
11 (benign and atypical) cases in order to assess the presence of the 24 studied criteria and to
12 determine the specificity and sensitivity (Table 2) of each criterion and the inter-observer
13 agreements for the 24 cytological studied criteria.

14 For this second reading AB and DG were blinded to the final diagnosis, and a delay of three
15 months was respected between the first and second reading.

16 A third reading was performed three months later by the experienced cytopathologist (DG) on
17 31 samples from the 135 cases, selected at random to evaluate the intra-observer agreement
18 on the criteria retained for the diagnostic tree.

19 ***Statistical analysis***

20 The Cohen’s kappa coefficient (k)¹³ was computed to assess inter-observer and intra-observer
21 agreement for the 24 cytological criteria. Generally, a k value less than 0.4 implies poor
22 agreement. A k value between 0.4 and 0.75 corresponds to moderate to good agreement while
23 a k value higher than 0.75 indicates excellent agreement.¹⁴

24 Sensitivity and specificity of the 24 cytological criteria were computed as well as their 95%
25 confidence intervals (95% CIs) using the Wilson method.

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3 1 We developed a score including the four non-100% sensitive and non-100% specific criteria
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5 2 harboring the highest specificities in order to improve their diagnostic performance by
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8 3 combining them. A logistic regression was used to assess if these criteria were associated
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10 4 with malignancy. A scoring system was then developed considering the values of the
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13 5 regression coefficients obtained from the logistic regression.

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15 6 XLSAT and STATA 16.1 were used for the analyses.

17 7 **RESULTS**

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20 8 *Inter-observer agreements between novice and experienced cytopathologists in biliary*
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22 9 *cytology* (Table 1).

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24 10 From the 24 studied cytological criteria the first reading highlighted 4 criteria with an
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26 11 excellent kappa value: “pigment rich background, bloody background, anisonucleosis,
27
28 12 irregular nuclear shape” and 6 criteria with a good kappa value: “inflammatory background,
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30 13 acinar arrangement, single malignant cell, N/C ratio > 0.5, clear chromatin and enhanced
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32 14 cytoplasmic membrane”.

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35 15 The second reading, performed after the elaboration of the learning atlas highlighted 14 other
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37 16 criteria which became highly reproducible with an excellent k value corresponding to the
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39 17 following criteria: “clean background, inflammatory background, epithelial cell density,
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41 18 nuclear overlap, 3D-cluster, single malignant cell, enhanced nuclear membrane, N/C ratio >
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43 19 0.5, clear chromatin, coarse chromatin, hyperchromatism, cytoplasmic vacuole, enhanced
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45 20 cytoplasmic membrane and neutrophils phagocytosis”. Moreover 4 criteria presented a good
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47 21 agreement: “necrotic background, mucinous background, acinar arrangement, and prominent
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49 22 nucleoli”. Kappa indices agreements of all the 24 criteria improved between the two readings
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52 23 and in the second reading 75% of them (18/24) were excellent.
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3 1 These results show the interest of the atlas for our training purpose. Indeed, we demonstrated
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5 2 that this atlas can represent a learning support for cytopathologists novices in biliary cytology.
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7 3 It highly improves the agreement between cytopathologists by providing a clear definition and
8
9 4 illustration of the cytological criteria.

5 ***Specificity and sensitivity of cytological criteria for the diagnosis of malignancy (Table 2)***

6 To reduce the number of retained cytological criteria, three criteria (clear, coarse chromatin
7
8 and hyperchromatism) were pooled into a single criterion entitled chromatin changes, as the
9
10 inter-observer agreements were excellent ($\kappa \geq 0.84$).

11 The frequency and the percentage of all the criteria were reported in 3 diagnostic groups:
12
13 malignant, atypical and benign cases. In addition, the sensitivity and specificity of each
14
15 criterion were analyzed in the malignant and non-malignant cases.

16 The only one 100% specific criterion was represented by the presence of “single malignant
17
18 cell”.

19 Three 100% sensitive criteria (irregular nuclear shape, chromatin changes and the presence of
20
21 cytoplasmic vacuole), were found in all malignant cases. This was not surprising as their
22
23 presence is linked to the definition of malignant cells (isolated or clustered).

24 Four other criteria harbored high specificities (between 89-98%): “3D-cluster, anisonucleosis
25
26 ($\geq 1:4$), N/C ratio > 0.5 and enhanced nuclear membrane”.

27 All the remaining studied criteria with low sensitivity ($< 49\%$) and/or a specificity less than
28
29 89% were not retained for further analysis.

30 ***Scoring system (Tables 3A and 3B)***

31 The four non-100% sensitive and non-100% specific criteria harboring the highest
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33 specificities (between 89-98%) were also very reproducible ($k \geq 0.88$) and were chosen to
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35 develop a scoring system of biliary malignancy. Each 4 criterion was pondered in a logistic
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37 regression analysis and a score was attributed based on their regression coefficients (Table
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3 1 3A). A final score resulted from the sum of each criterion's score. Table 3B reports the
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5 2 sensitivities and specificities associated with each value of the final score. A final score > 16
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7 3 was associated with the best specificity for malignancy, and more specifically, the presence of
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9 4 3 criteria, namely "3D-cluster, anisonucleosis and N/C ratio > 0.5", allowed to reach a
10
11 5 specificity of 100% (95% CI: 94-100%) and a sensitivity of 72% (95% CI: 61-82%) for the
12
13 6 diagnosis of adenocarcinoma. As shown in table 3A, the criterion "enhanced nuclear
14
15 7 membrane" which showed the lowest score appeared less essential for the accurate diagnosis
16
17 8 and wasn't retained for the diagnostic tree.

9 ***Malignant diagnostic tree (Fig. 2)***

10 The main goal of this diagnostic tree was to avoid false positive results either by identifying a
11
12 11 single malignant cell (the only 100% specific criterion) or by summing the following 3
13
14 12 criteria: "3D-cluster, anisonucleosis ($\geq 1:4$) and N/C ratio > 0.5", reaching a 100% specificity
15
16 13 (final score > 16).

17 The intra-observer agreement of the 4 selected criteria included in this diagnostic tree was
18
19 14 assessed by a third reading conducted by the experienced cytopathologist (DG) blinded to the
20
21 15 final diagnosis on 32 biliary samples randomly selected among the 135 cases. All the 4
22
23 16 criteria were highly reproducible with excellent intra-observer kappa indices (Table 4).

24 The diagnostic tree was then applied to the second reading of both the experienced
25
26 18 cytopathologist (DG) and the novice cytopathologist in biliary cytology (AB) in order to
27
28 19 assess its internal validity on the 135 samples. No false positive or false negative results were
29
30 20 obtained when the diagnostic tree was applied to the experienced pathologist's second
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32 21 reading. When applying it to the novice's second reading, both diagnostic specificity and
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34 22 sensitivity were 97%. Indeed, there were 3 false negatives (3 well-differentiated
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36 23 adenocarcinomas) and one false positive (PSC).

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1 DISCUSSION

2 Our study is the first to propose an original diagnostic tree reaching a 100% specificity in a
3 large population of well-differentiated adenocarcinomas regardless of the reader's experience
4 in biliary cytology. We also produce a bile cytology learning atlas improving intra- and inter-
5 observer agreements for the training of cytopathologists novices in biliary cytology.

6 The main goal of our study was to determine the most specific, sensitive and reproducible
7 cytodiagnostic criteria for the diagnosis of adenocarcinomas, especially the well-differentiated
8 ones which represent a diagnostic challenge.

9 We first studied a scoring system based on 4 specific, sensitive and reproducible criteria for
10 malignancy (“3D-cluster, anisonucleosis, N/C ratio > 0.5 and enhanced nuclear membrane”).

11 When bile specimens harbored two of them with a score higher than 10, the specificity
12 reached 97% and the sensitivity of diagnosis of malignancy was 96%. As we focused on the
13 specificity rather than the sensitivity in order to avoid false positives, we built our diagnostic
14 tree with the more efficient (specific and reproducible) cytological criteria of malignancy. The
15 decisive diagnostic tree encompasses the only 100% specific criterion (single malignant cell)
16 and three out of the four scoring system’s criteria (“3D-cluster, anisonucleosis and enhanced
17 nuclear membrane”). Indeed the “enhanced nuclear membrane” criterion was not retained for
18 the elaboration of the diagnostic tree as it was not seen by the MGG staining and as its
19 individual score and sensitivity were the lowest of the 4 scoring system’s criteria.

20 Our study highlighted three 100% sensitive criteria named “chromatin changes, vacuolated
21 cytoplasm and irregular nuclear shape”. In other words, they are sufficient to affirm the
22 malignancy and represent strong criteria in our series including 81% of well-differentiated
23 adenocarcinomas compared to 2 other previous studies reporting only 43% and 63% of well-
24 differentiated adenocarcinomas respectively.^{12,15}

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3 1 To our knowledge only 3 multivariate studies performed on biliary cytology reported a
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5 2 scoring system encompassing 4 or 6 criteria for malignancy in the literature.^{12,15,16}
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8 3 Unlike our study, their scoring systems were established by experienced cytopathologists. The
9
10 4 more recent one (2019) conducted in Japan was based on 89 cholangiocarcinomas.¹² The
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12 5 presence of 3 out of 4 criteria (abnormal chromatin, irregularly arranged nuclei, irregularly
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14 6 overlapped nuclei and irregular cluster margins) reached a diagnostic specificity of 98% and a
15
16 7 sensitivity of 87%. The second study by Jin YH *et al.* was performed on a cohort of 43
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18 8 adenocarcinomas and showed that the presence of 3 out of the 4 following criteria
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20 9 (hyperchromatism, coarse chromatin, increased N/C ratio, loss of honeycomb arrangement)
21
22 10 reached a diagnostic specificity of 90% and a sensitivity of 65%. Finally, another Japanese
23
24 11 team reported in a small cohort of 22 adenocarcinomas a scoring system based on 6 criteria
25
26 12 (loss of honeycomb arrangement, enlarged nuclei, loss of polarity, bloody background, flat
27
28 13 nuclei and cell-in-cell arrangement). When more than 3 out of the 6 criteria were found, the
29
30 14 specificity reached 76.9% and the overall sensitivity of diagnosis of malignancy was 86.4%.¹⁵
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33 15 Two out of the 4 final retained criteria for establishing our proposed diagnostic tree were
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35 16 already reported in these three bile cytology studies even if their terminology of the criteria
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37 17 and/or their definition differs somewhat: “N/C ratio > 0.5” by Jin et al. (increased N/C ratio)
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39 18 and Nakajime et al. (named enlarged nuclei) and “3D-cluster” criterion combines some criteria
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41 19 of the 3 studies (loss of honeycomb arrangement, irregularly arranged nuclei, irregularly
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43 20 overlapped nuclei and loss of polarity). The criteria “single malignant cell” and
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45 21 “anisonucleosis” were not retained in their scoring system.
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51 22 Moreover, our diagnostic tree performed well whatever the cytopathologist’s experience in
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53 23 biliary cytology. Applying the diagnostic tree to the experienced cytopathologist's second
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55 24 reading, no false positive or false negative were noted. When applying it to the novice's
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57 25 second reading, both diagnostic specificity and sensitivity were 97%, better than the 3
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1 multivariate studies performed by experienced cytopathologists. The only false positive was
2 observed in a context of PSC, a classic pitfall. It should be noted that the 2nd reading was
3 blind to any clinical information. That is why some authors have already pointed out the need
4 of a multidisciplinary management not to ignore this diagnostic trap.^{8,17-19} In the specific
5 context of PSC or stents, Goyal et al. even recommend an anisonucleosis > 1:6 to avoid false
6 positives.¹⁷

7 Three false negative results were also noted, all corresponding to well-differentiated
8 adenocarcinomas where the “single malignant cell” criterion had not been found and less than
9 three criteria had been identified. Furthermore it is interesting to note that each case included
10 8 bile cytology slides. It is commonly accepted that increasing the number of slides to be read
11 reduces the cytopathologist's concentration and therefore increases the risk of missing single
12 malignant cells.

13 Since bile cytology is an uncommon and not widely taught cytology, our second objective
14 was to produce a learning atlas to increase the inter-observer agreements between
15 cytopathologists and to train cytopathologists novices in biliary cytology. It can be noted that
16 the novice's results are good from the first reading, which can be explained by the fact that the
17 studied-criteria corresponded to “general” cytological criteria and that the novice had
18 experience in cytology. Our study demonstrates that by redefining the less reproducible
19 criteria and by elaborating an atlas illustrating each studied cytological criterion, all the inter-
20 observer agreements improved between the first and the second readings and 75% of them
21 became excellent. These results represent another strength of our study since none of the three
22 multivariate analyses performed on bile specimens investigated the reproducibility of the
23 selected criteria.^{12,15,16}

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3 1 MGG staining did not allow the assessment of three criteria, namely “enhanced nuclear
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5 2 membrane, prominent nucleoli and chromatin changes” pointing the superiority of
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7 3 Papanicolaou staining for biliary samples.
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10 4 To conclude, our diagnostic tree of malignancy on bile aspiration specimens is based on 4
11
12 5 efficient criteria of well-differentiated adenocarcinoma which allow together with a
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14 6 multidisciplinary approach, a diagnosis of adenocarcinoma with a specificity of 100% and a
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16 7 high sensitivity. It comes with a learning atlas useful for the training and accuracy of
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18 8 cytopathologists. This atlas could also be the subject of data integration in artificial
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20 9 intelligence. Indeed, the precise definitions and illustrations of the cytological criteria defined
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22 10 in our atlas, could be used as a support for learning an artificial intelligence software
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24 11 dedicated to the screening of biliary adenocarcinomas, providing a real help to
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26 12 cytopathologists, using scanners able to analyze 3D images, especially 3D-clusters as part of
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28 13 our diagnostic tree.
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32 33 **REFERENCES**

- 34
35 15 1. Bouvier A-M, Remontet L, Jouglu E, Launoy G, Grosclaude P, Buémi A, *et al.* Incidence of
36
37 16 gastrointestinal cancers in France. *Gastroenterol Clin Biol.* oct 2004;28:877-81.
38
39 17 2. Cowppli-Bony A, Colonna M, Ligier K, Jooste V, Defossez G, Monnereau A, *et al.*
40
41 18 *Épidémiologie descriptive des cancers en France métropolitaine: incidence, survie et*
42
43 19 *prévalence. Bull Cancer (Paris).* July 2019;106 (7-8):617-34.
44
45 20 3. Roth GS, Bichard P, Fior-Gozlan M, *et al.* Performance of bile aspiration plus brushing to
46
47 21 diagnose malignant biliary strictures during endoscopic retrograde cholangiopancreatography.
48
49 22 *Endosc Int Open.* sept 2016;4(9):E997-1003.
50
51
52 23 4. Fior-Gozlan M, Giovannini D, Rabeyrin M, Mc Leer-Florin A, Laverrière M-H, Bichard P.
53
54 24 *Monocentric study of bile aspiration associated with biliary brushing performed during*
55
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2
3 1 endoscopic retrograde cholangiopancreatography in 239 patients with symptomatic biliary
4
5 2 stricture. *Cancer Cytopathol.* May 2016;124(5):330-9.
- 6
7 3 5.Kurzawinski TR, Deery A, Dooley JS, Dick R, Hobbs KE, Davidson BR. A prospective
8
9 4 study of biliary cytology in 100 patients with bile duct strictures. *Hepato*l Baltim Md. déc
10
11 5 1993;18(6):1399-403.
- 12
13 6 6.Fogel EL, Sherman S. How to Improve the Accuracy of Diagnosis of Malignant Biliary
14
15 7 Strictures. *Endoscopy.* Nov 1999;31(9):758-60.
- 16
17 8 7.Pitman MB, Centeno BA, Ali SZ, *et al.* Standardized terminology and nomenclature for
18
19 9 pancreatobiliary cytology: the Papanicolaou Society of Cytopathology guidelines. *Diagn*
20
21 10 *Cytopathol.* April 2014;42(4):338-50.
- 22
23 11 8.Furmanczyk PS, Grieco VS, Agoff SN. Biliary brush cytology and the detection of
24
25 12 cholangiocarcinoma in primary sclerosing cholangitis: evaluation of specific cytomorphologic
26
27 13 features and CA19-9 levels. *Am J Clin Pathol.* Sept 2005;124(3):355-60.
- 28
29 14 9.Geramizadeh B, Moughali M, Shadim-Aein A *et al.* False negative and false positive rates
30
31 15 in common bile duct brushing cytology, a single center experience. *Gastroenterol Hepatol*
32
33 16 *Bed Bench.* 2018;11(4):296-300.
- 34
35 17 10.Avadhani V, Hacıhasanoglu E, Memis B, *et al.* Cytologic predictors of malignancy in bile
36
37 18 duct brushings: a multi-reviewer analysis of 60 cases. *Mod Pathol Off J U S Can Acad Pathol*
38
39 19 *Inc.* 2017;30(9):1273-86.
- 40
41 20 11. Barr Fritcher EG, Caudill JL, Blue JE, *et al.* Identification of malignant cytologic criteria
42
43 21 in pancreatobiliary brushings with corresponding positive fluorescence in situ hybridization
44
45 22 results. *Am J Clin Pathol.* Sept 2011;136(3):442-9.
- 46
47 23 12. Hayakawa C, Hoshikawa M, Imura J, Ueno T, Koike J. Bile cytology: A new scoring
48
49 24 system for improving diagnostic accuracy. *Diagn Cytopathol.* July 2019;47(7):641-7.
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3 1 13. Cohen J. Weighted kappa: nominal scale agreement with provision for scaled
4
5 2 disagreement or partial credit. *Psychol Bull.* Oct 1968;70(4):213-20.
6
7 3 14. Fleiss J. *Statistical Methods for Rates and Proportions*, 2nd ed. New York: John Wiley
8
9 4 and Sons; 1981.
10
11 5 15. Nakajima T, Tajima Y, Sugano I, *et al.* Multivariate statistical analysis of bile cytology.
12
13 6 *Acta Cytol.* Febr 1994;38(1):51-5.
14
15 7 16. Jin YH, Kim SH, Park CK. Diagnostic criteria for malignancy in bile cytology and its
16
17 8 usefulness. *J Korean Med Sci.* Dec 1999;14(6):643-7.
18
19 9 17. Goyal A, Sharaiha RZ, Alperstein SA, Siddiqui MT. Cytologic diagnosis of
20
21 10 adenocarcinoma on bile duct brushings in the presence of stent associated changes: A
22
23 11 retrospective analysis. *Diagn Cytopathol.* Oct 2018;46(10):826-32.
24
25 12 18. Geramizadeh B, Moughali M, Shahim-Aein A, *et al.* False negative and false positive
26
27 13 rates in common bile duct brushing cytology, a single center experience. *Gastroenterol*
28
29 14 *Hepatol Bed Bench.* 2018;11(4):296-300.
30
31 15 19. Salomao M, Gonda TA, Margolskee E *et al.* Strategies for improving diagnostic accuracy
32
33 16 of biliary strictures. *Cancer Cytopathol.* 2015 Apr;123(4):244-52.
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42 **Table legends**

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44 19 Table 1. Inter-observer agreements between a novice (AB) and an experienced
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46 20 cytopathologist (DG) in biliary cytology for the 24 criteria in the 69 malignant cases: first and
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48 21 second readings.

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50 22 Table 2. Each criterion used with its frequency and percentage (%) in malignant and non-
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52 23 malignant cases (atypical and benign), their sensitivity, specificity with their 95% confidence
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54 24 intervals (CIs 95%).

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56 25 Table 3.A. Scoring system.
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3 1 Table 3.B. Diagnostic performance of the combination of the retained criteria.
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5 2 Table 4. Kappa intra-observer indices (DG) of the 4 criteria of the malignant diagnostic tree
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10 4 **Figure legends**
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12 5 Figure 1. Bile cytology learning atlas illustrating the studied cytological criteria. PAPA:
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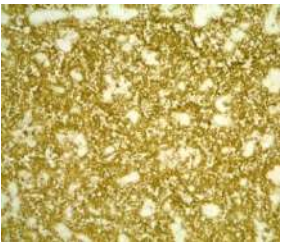
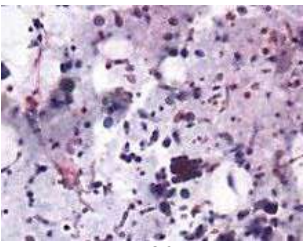
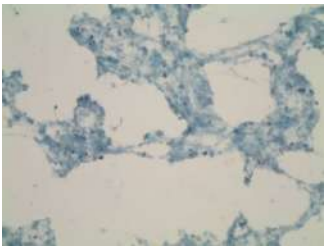
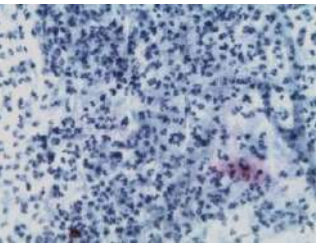
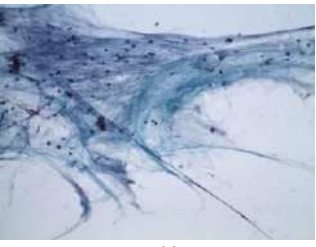

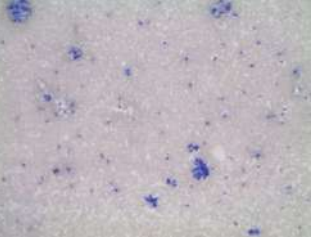
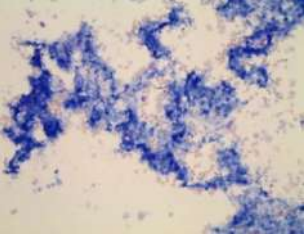
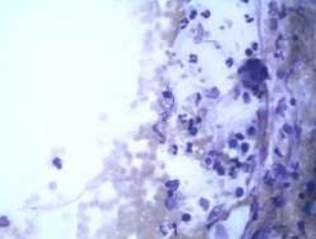
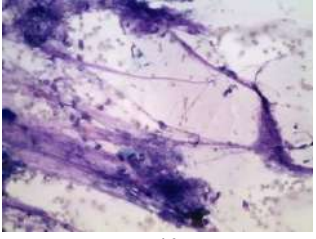
14 6 Papanicolaou stain, MGG: May Grünewald Giemsa stains.
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17 7 Figure 2. Diagnostic tree to assert the diagnosis of adenocarcinoma on bile aspiration
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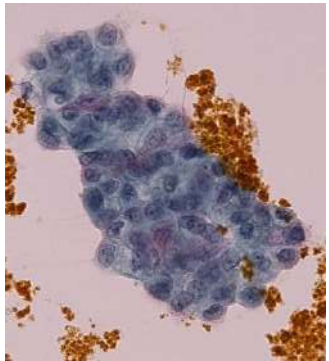
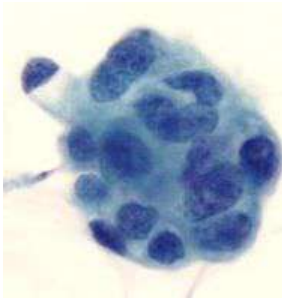

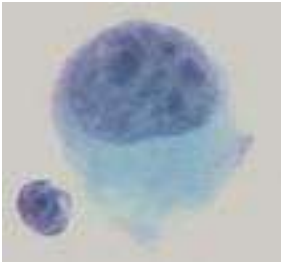
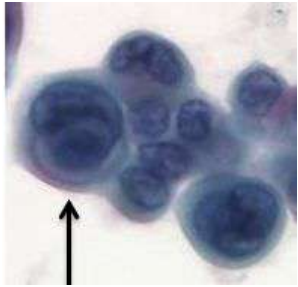
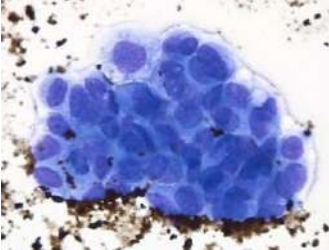
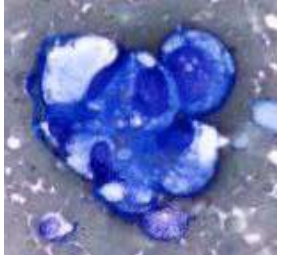
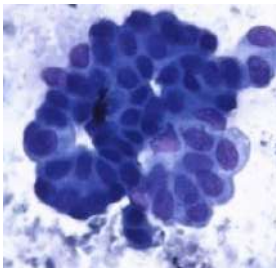

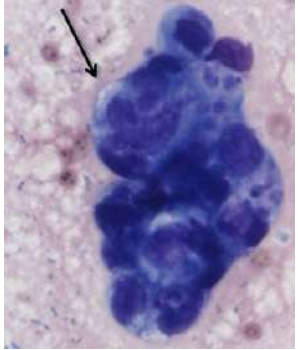
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BACKGROUND OF BILE CYTOLOGY

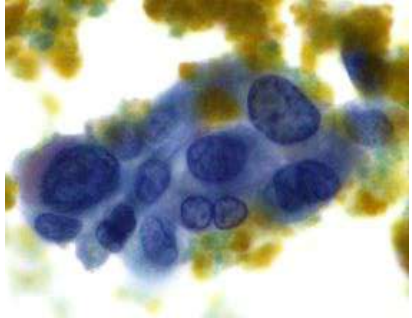
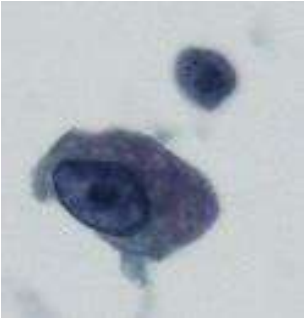
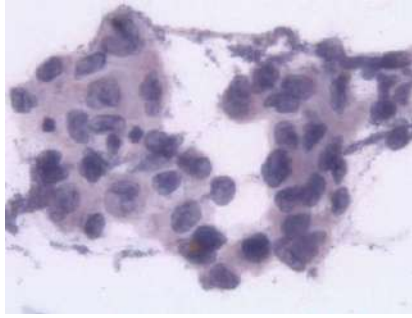
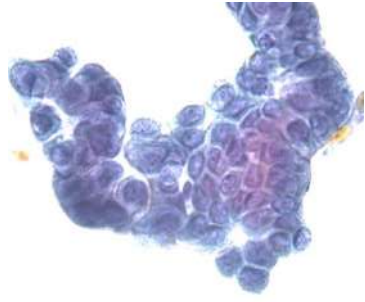
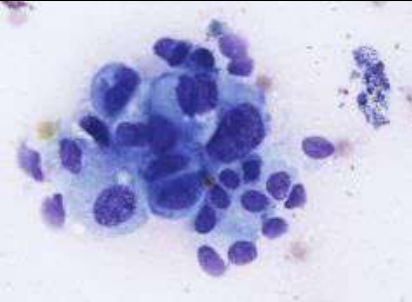

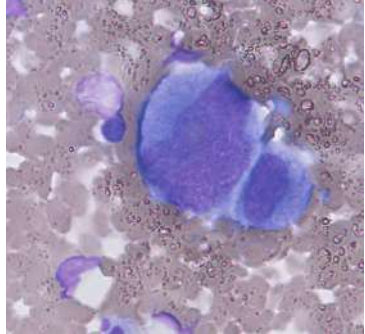
	Pigment-rich	Bloody	Necrotic	Inflammatory	Mucinous
PAPA	 x 5	 x 20	 x 20	 x 40	 x 40
MGG	 x 5	 x 5	 x 20	 x 40	 x 40

PRESENTATION OF EPITHELIAL CELLS OR CLUSTERS

	<p>Nuclear overlap Disorganized arrangement of nuclei in rows and columns with loss of polarity</p>	<p>3D-cluster Tightly cohesive cell balls with considerable nuclear overlapping, high N/C ratio</p>	<p>Acinar arrangement</p>	<p>Single malignant cell Single cell with intact cytoplasm, nuclei occupying at least 50% of the cell area, chromatin changes, irregular nuclear shape with or without prominent nucleoli</p>	<p>Cytophagy Epithelial cell-in-cell arrangement in single or cell cluster</p>
<p>PAPA</p>	 <p>x 40</p>	 <p>x 40</p>	 <p>x 40</p>	 <p>x 63</p>	 <p>x 40</p>
<p>MGG</p>	 <p>x 40</p>	 <p>x 40</p>	 <p>x 40</p>	 <p>x 63</p>	 <p>x 40</p>


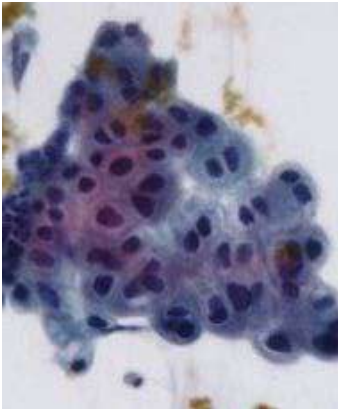
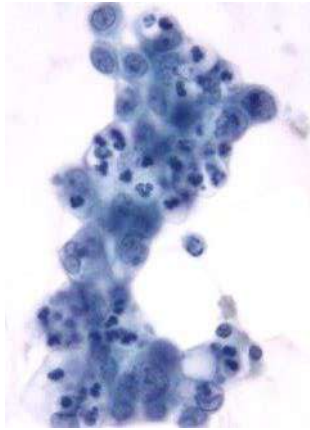
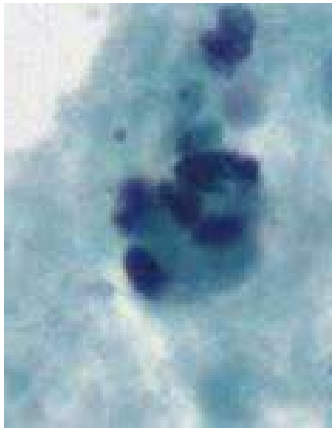
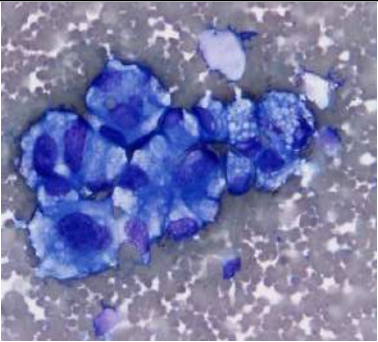
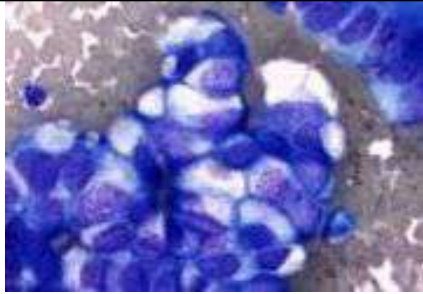
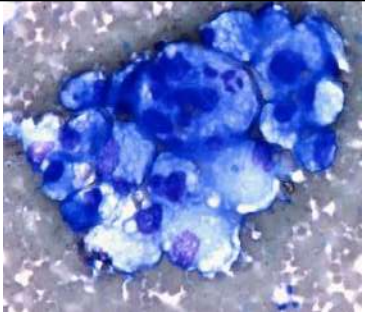

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PRESENTATION OF EPITHELIAL CELLS OR CLUSTERS

	Anisonucleosis ($\geq 1:4$) Size variation with at least 4-fold variation in nuclear size	Enhanced nuclear membrane Visible at 20 x objective lens	Irregular nuclear shape Including nuclear grooves and flat nuclei	N/C ratio > 0.5
PAPA	 x 40	 x 40	 x 40	 x 40
MGG	 x 40	Not seen with MGG staining	 x 40	 x 63

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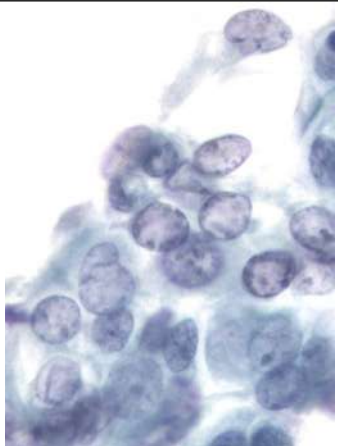
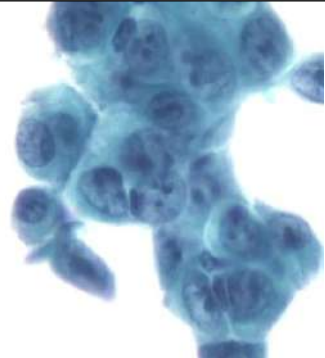
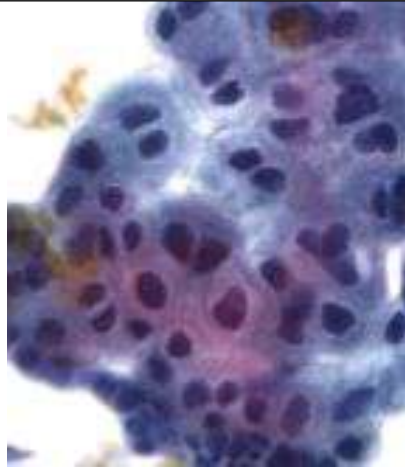
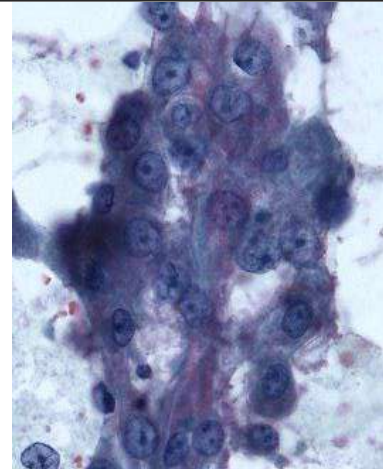
PRESENTATION OF EPITHELIAL CELLS OR CLUSTERS

	Vacuolated cytoplasm	Enhanced cytoplasmic membrane	Neutrophils phagocytosis	Degenerative cell
PAPA	 <p>x 63</p>	 <p>x 40</p>	 <p>x 40</p>	 <p>x 40</p>
MGG	 <p>x 40</p>	 <p>x 40</p>	 <p>x 63</p>	 <p>x 40</p>

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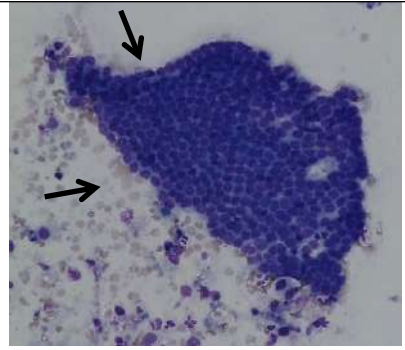
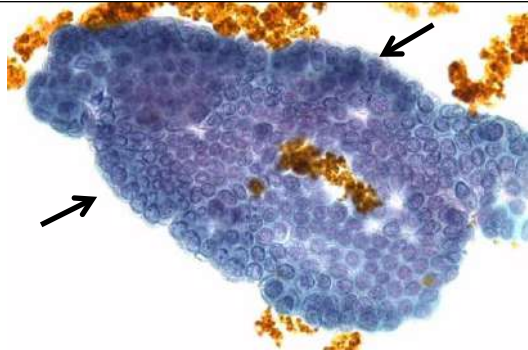
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CHROMATIN CHANGES CRITERIA (Not seen with MGG staining)

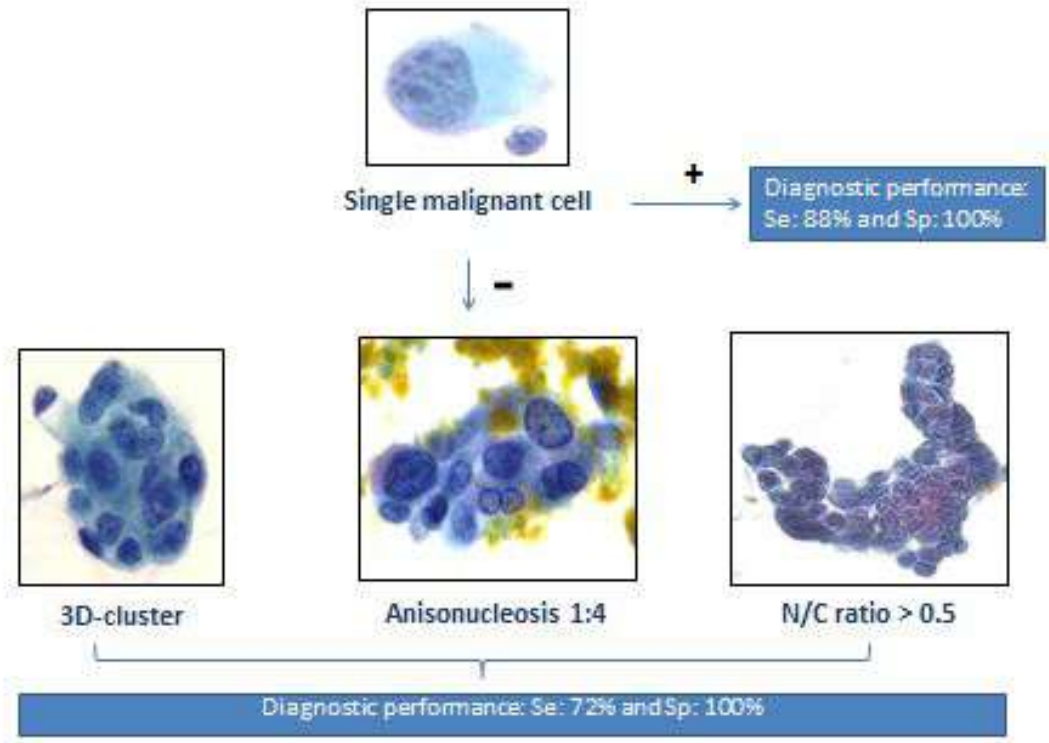
	Clear chromatin	Coarse chromatin	Hyperchromatism	Prominent nucleoli
PAPA	 x 40	 x 40	 x 40	 x 40

NORMAL BILIARY FLAPS

All the previous cytological criteria must be compared to the nuclei of the benign ductal epithelium stained by PAPA (x 40 objective lens) and/or MGG (x 20 objective lens) characterised by large, flat, monolayered cell sheet well-limited by a palissadic border (arrows). The cells are mostly cohesive, with well-defined cell borders and polarity. They possess small, centrally placed, round oval, regular nuclei and abundant cytoplasm. The chromatin is fine and nucleoli inconspicuous.



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Categories	Criteria	First reading k (95% CI)	Second reading k (95% CI)
1-Background	Pigment-rich	0.97 (0.91-1)	1 (1-1)
	Clean	0.54 (0.34 - 0.73)	0.88 (0.8-0.97)
	Bloody	0.84 (0.7-0.99)	0.95 (0.89-1)
	Necrotic	0.58 (0.24-0.91)	0.64 (0.4-0.88)
	Inflammatory	0.66 (0.45-0.86)	0.92 (0.84-1)
	Mucinous	0.33 (0.15-0.5)	0.64 (0.49-0.79)
2-Presentation	Cell density	0.35 (0.19 - 0.52).	0.86 (0.79-0.94)
	Nuclear overlap	0.39 (0.15-0.92)	1 (1-1)
	3D-cluster	0.43 (0.26-0.61)	0.88 (0.79-0.96)
	Acinar arrangement	0.64 (0.46-0.82)	0.69 (0.57-0.81)
	Single malignant cell	0.68 (0.49-0.86)	0.95 (0.9-1)
	Cytophagy	0.12 (0.095-0.33)	0.3 (0.094-0.51)
3-Nuclei	Anisonucleosis	0.90 (0,71-1)	1 (1-1)
	Irregular nuclear shape	0.99 (0.93-1)	1 (1-1)
	Enhanced nuclear membrane	0.31 (0.12-0.5)	0.89 (0.81-0.98)
	N/C ratio > 0.5	0.67 (0.51 – 0.83)	0.99 (0.97-1)
	Clear chromatin	0.74 (0.51-0.98)	0.96 (0.91-1)
	Coarse chromatin	0.18 (0.007-0.35)	0.84 (0.75-0.94)
	Hyperchromatism	0.54 (0.36-0.72)	0.92 (0.82-1)
	Prominent nucleoli	0.18 (0.007-0.35)	0.73 (0.64-0.83)
4-Cytoplasm	Vacuolated cytoplasm	0.54 (0.36-0.72)	0.97 (0.92-1)
	Enhanced cytoplasmic membrane	0.63 (0.48 - 0.79)	0.94 (0.88-1)
5-Accompanying criteria	Neutrophils phagocytosis	0.31 (0.19-0.8)	0.84 (0.73-0.95)
	Degenerative cell	0.25 (0.05-0.44)	0.48 (0.25-0.71)

Categories	Criteria	Malignant (N=69)	Non-Malignant		Sensitivity (CIs 95%)	Specificity (CIs 95%)
			Atypical (N=11)	Benign (N=55)		
1-Background	Pigment-rich	46 (66%)	5 (45%)	33 (60%)	65% (53-78%)	41% (33-49%)
	Clean	23 (33%)	3 (27%)	19 (34%)	41% (28-53%)	74% (67-82%)
	Bloody	16 (23%)	2 (18%)	10 (21%)	26% (14-38%)	85% (77-93%)
	Necrotic	9 (13%)	1 (9%)	4 (7%)	10% (0-22%)	89% (81-97%)
	Inflammatory	14 (20%)	4 (36%)	14 (25%)	23% (11-35%)	76% (68-83%)
	Mucinous	17 (24%)	5 (45%)	19 (34%)	16% (4-28%)	55% (47-62%)
2-Presentation	Cell density	34 (50%)	4 (36%)	18 (33%)	84% (77-89%)	30% (23-38%)
	Nuclear overlap	67 (97%)	5 (45%)	5 (9%)	97% (93-99%)	79% (71-85%)
	3D-cluster	55 (80%)	1 (9%)	0	79% (71-85%)	98% (94-99%)
	Acinar arrangement	59 (86%)	0	1 (2%)	85% (78-90%)	98% (94-99%)
	Single malignant cell	61 (88%)	0	0	88% (81-92%)	100% (97-100%)
	Cytophagy	24 (35%)	0	0	35% (27-43%)	100% (97-100%)
	Anisonucleosis	62 (90%)	2 (18%)	0	89% (82-93%)	97% (92-99%)
3-Nuclei	Irregular nuclear shape	69 (100%)	7 (64%)	16 (29%)	100% (97-100%)	62% (54-70%)
	Enhanced nuclear membrane	41 (59%)	2 (18%)	0	59% (54-67%)	97% (93-99%)
	N/C ratio > 0.5	68 (98%)	5 (45%)	2 (4%)	98% (94-99%)	89% (83-93%)
	Clear chromatin	45 (65%)	6 (54%)	19 (34%)	86% (73-98%)	83% (76-91%)
	Coarse chromatin	35 (50%)	3 (27%)	14 (25%)	65% (53-78%)	89% (82-97%)
	Hyperchromatism	17 (24%)	0	5 (9%)	30% (18-43%)	98% (91-100%)
	Chromatin changes	69 (100%)	6 (54%)	10 (18%)	100% (97-100%)	75% (67-81%)
	Prominent nucleoli	55 (80%)	5 (45%)	11 (20%)	94% (89-97%)	56% (48-64%)
4-Cytoplasm	Vacuolated cytoplasm	69 (100%)	5 (45%)	16 (29%)	100% (97-100%)	68% (60-75%)
	Enhanced cytoplasmic membrane	34 (49%)	5 (45%)	14 (25%)	49% (41-57%)	71% (63-78%)
5-Accompanying criteria	Neutrophils phagocytosis	25 (36%)	3 (27%)	6 (1%)	36% (28-44%)	86% (79-91%)
	Degenerative cell	17 (25%)	1 (9%)	2 (3%)	25% (13-37%)	95% (87-100%)

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Criteria	Regression coefficients	Score
3D-cluster	5.54 (3.48 – 7.60)	6
Anisonucleosis	5.65 (4.04 – 7.26)	6
N/C ratio > 0.5	6.35 (4.23 – 8.48)	6
Enhanced nuclear membrane	3.85 (2.36 – 5.33)	4

Final score	Specificity (95% CIs)	Sensitivity (95% CIs)
> 6	95% (87-98%)	96% (88-99%)
> 10	97% (90-99%)	96% (88-99%)
> 12	98% (92-100%)	86% (75-92%)
> 16	100% (94-100%)	72% (61-82%)

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Four retained criteria	Third reading k (95% CIs)
Single malignant cell	0,87 (0,7-1)
3D-cluster	0.80 (0,6-1)
Anisonucleosis (1:4)	0.94 (0,81-1)
N/C ratio > 0.5	0.87 (0,69-1)